# Determination of Free (Unbound) Anamorelin in Human Plasma by Ultrafiltration

Gene Ray<sup>1</sup>, Elle deGroot<sup>2</sup>, Yu-Hui (Ann) Fu<sup>1</sup>, Matalin Shine<sup>1</sup> and Yansheng Liu<sup>1</sup>

<sup>1</sup>KCAS (Shawnee, Kansas) and <sup>2</sup> Helsinn Therapeutics (Bridgewater, New Jersey)



# INTRODUCTION

Anamorelin is under investigation for cancer cachexia, a devastating, often late-stage complication of an underlying malignancy. Despite the significant importance of cancer-related cachexia, treatments are lacking and no products are approved for this indication. Anamorelin, by virtue of its ghrelin agonist activity, may serve a role in the treatment of cancer cachexia.

To evaluate the extent of free (unbound) Anamorelin in human plasma, the appropriate procedures for processing samples were investigated. Ultrafiltration was found to be valid technique with limited non-specific binding (NSB) observed to the device. LC-MS/MS methods were then validated for the analysis of free and total Anamorelin.

$$H_3C$$
 $H_2N$ 
 $CH_3$ 
 $CH_3$ 
 $CH_3$ 
 $CH_3$ 

**Chemical Structure** 

# METHOD

#### **Protein Binding Assessments**

- Non-Specific Binding
- Concentration Dependency
- Affect of pH
- Affect of Freeze/Thaw
- Anticoagulant (Heparin vs. EDTA)
- Binding to Selected Proteins

#### **Ultrafiltration**

Device: Amicon Centrifree®, YM-30 Centrifugation: 1000 x g, Fixed Angle Rotor Maximum Filtration: 1/3 Sample Volume

Temperature: 37 ± 2 °C

#### Sample Extraction

Plasma (50 mcL) or Ultrafiltrate (150 mcL) were combined with 400 mcL internal standard (Piroxicam in MeOH/ACN, 50:50). Samples were centrifuged 5 min at 15,000 rpm and the resulting supernatants combined 1:1 with 10 mM ammonium formate buffer, pH 3.2, and analyzed by LC-MS/MS.

# METHOD (Cont.)

Extracts were analyzed using fully validated methods for Human Plasma (5 to 1280 ng/mL)
Human Plasma Ultrafiltrate (1 to 100 ng/mL)

## **Liquid Chromatography**

HPLC : Shimadzu 10AD
Autosampler : Perkin Elmer 200
Column : Pursuit C18 (50 x 2 mm) 5μ

Temperature: 30 °C

MP A: ACN/Ammonium Formate Buffer, pH 3.2 (10/90)

MP B: ACN/MeOH (40/60) Flow Rate: 0.35 mL/min Rinse: ACN/DI H<sub>2</sub>O (90/10)

## **Mass Spectrometry**

AB Sciex API 3000 with Turbo-ion Spray interface

Polarity : Positive

Scan type: Multiple Reaction Monitoring

Anamorelin:  $547.5 \rightarrow 276.3 \ m/z$ Piroxicam (IS):  $332.0 \rightarrow 164.0 \ m/z$ 

## RESULTS

#### Non-Specific Binding (NSB)

Ultrafiltrate <u>Conc.</u> ng/mL	Recovery %	NSB %	
3	92.4	7.6	
15	93.9	6.1	

#### **Concentration Dependency**

Plasma <u>Conc.</u> ng/mL	Free Fraction (Mean) %
100 200	$3.57 \pm 0.27$ $3.59 \pm 0.39$
300	$3.91 \pm 0.35$
375	$3.67 \pm 0.40$
750	$4.05 \pm 0.21$
1000	$4.43 \pm 0.27$
1500	$4.66 \pm 0.12$

Li Heparin, n = 5 replicates, pH 7.40 at 37 °C

# RESULTS (Cont.)

## Affect of pH

Free Fraction (Mean) %	
$6.91 \pm 0.11$ $6.54 \pm 0.34$ $5.80 \pm 0.13$	

Li Heparin, n = 5 replicates, c = 375 ng/mL at 37 °C. Plasma pH was adjusted using carbon dioxide gas.

#### Affect of Freeze/Thaw

Anamorelin <u>Conc.</u> ng/mL	Plasma	Free Fraction (Mean) %
200		4.63 ± 0.14
200	Fresh	7.00 ± 0.17
375	(Non-Frozen)	5.23 ± 0.01
200	3 Cycles	4.86 ± 0.12
375	Freeze/Thaw	4.93 ± 0.07

Li Heparin, n = 4 replicates at pH 7.40 at 37 °C.

### Affect of Anticoagulant

Slightly lower free fractions were observed for Anamorelin in plasma containing Li heparin (3.57 – 4.66%) compared to plasma containing K<sub>2</sub>EDTA anticoagulant (4.06 – 5.15%).

#### **Binding to Selected Plasma Proteins**

Anamorelin <u>Conc.</u> ng/mL	Plasma Protein	Free Fraction (Mean) %
200	$\alpha_1$ –Acid Glycoprotein (AGP)	2.79
375		3.05
200	Human Albumin	62.8
375		63.7

AGP, 1.25 mg/mL and Albumin, 40 mg/mL in PBS, pH 7.40; (n = 4 replicates) at 37 °C.

# RESULTS (Cont.)

#### **Clinical Trial Results**

Protein binding assessment for Anamorelin have been conducted in three separate clinical trails.

**ST-ANAM-110** entitled "An Open-Label, Single and Multiple-Dose, Pharmacokinetic, Safety and Tolerability Study of Anamorelin HCl in Healthy Normal Volunteers".

RC-1291-203 entitled "A Two-Part Phase II Study of the Effects of RC-1291HCI in Patients with Cancer Anorexia/Cachexia: An In-patient Crossover Study of the Appetite-Enhancing and Biochemical Effects of RC-1291 HCI followed by an Out-patient Parallel Group Study of the Effects of RC-1291 HCI on Body Weight, Body Composition and Functional Performance".

**ST-ANAM-207** entitled "A Randomized, Double-Blind, Placebo-Controlled, Multicenter Phase II Anamorelin HCI Dose Range Study to Evaluate the Safety and Efficacy of Anamorelin HCI in Patients with NSCLC".

Subject		Free Fraction (%)		
<u>Status</u>	<u>Protocol</u>	<u>Mean</u>	Low	<u>High</u>
Healthy	ST-ANAM-110	6.31	3.86	10.6
Cancer	RC-1291-203	4.07	1.94	7.07
Cancer	ST-ANAM-207	4.53	2.09	8.61

## CONCLUSION

An accurate procedure was developed and validated for the determination of free Anamorelin in human plasma samples using ultrafiltration. Of significance was the high percentage to which Anamorelin binds to  $\alpha_1$ -acid glycoprotein ( $\geq 97\%$ ) with nominal binding to albumin ( $\sim 36.5\%$ ).

The results from clinical sample analyses suggests higher extent of plasma protein binding in subjects with cancer (4.1- 4.5% free) compared to healthy normal subjects (6.3% free). The upcoming investigation will now explore the correlation between the extent of Anamorelin protein binding and  $\alpha_1$ -acid glycoprotein levels.